Bullet Blender[®] 50 Homogenization Protocol for Shellfish

The protocol described in this document is for the use of the Bullet Blender[®] 50 for the homogenization of shellfish. This protocol was created using Mahogany clam tissue. Other types of shellfish (mussels, clams, scallops, etc.) may require a slightly modified homogenization protocol. This protocol does not specify a particular buffer - you may choose which is most appropriate for your downstream application (nucleic acid isolation, protein extraction, etc.).

Materials Required:

shellfish , Bullet Blender[®] 50, homogenization buffer, pipettor, 50mL centrifuge tubes, 4.8mm stainless steel beads (part number SSB48).

Instructions

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- 1. Break the shell of the animal in a clean dissection area. Remove fragments of shell from the soft tissue.
- 2. If desired, wash the soft tissue with saline to remove any sand or shell fragments.
- 3. Blot excess liquid from the soft tissue using a Kimwipe[®] or other lint free cloth.
- 4. Cut the tissue into appropriately sized pieces (up to 9g).
- 5. Place tissue into a 50mL skirted conical centrifuge tube (Axygen[®] or Corning[®]).
- 6. Add a mass of stainless steel beads (4.8mm) to the tube equal to approximately 3x the mass of your sample.
- 7. Add 4-7mL buffer.
- 8. Screw caps onto centrifuge tubes TIGHTLY.
- 9. Place tubes into the Bullet Blender[®] 50.
- 10. Set controls for SPEED 10 and TIME 12 minutes.
- 11. Remove tubes from the instrument.
- 12. Visually inspect samples, if homogenization is unsatisfactory, run for another six minutes at the SPEED 10.
- 13. Proceed with your downstream application.

SAFETY NOTE!!!

When using a centrifuge to separate your homogenate from the debris and beads, make sure your tubes are balanced.

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