Protocol for E. coli Cultures Homogenization in the Bullet Blender[®]

The protocol described in this document is for the use of the Bullet Blender[®] for the homogenization of Escherichia coli (or other bacterial) cultures. This protocol does not specify a particular buffer - you may choose which is most appropriate for your downstream application (nucleic acid isolation, protein extraction, etc.).

Materials Required:

E. coli, aspirator, Bullet Blender[®], homogenization buffer, pipettor, microcentrifuge tubes, and 0.1 mm glass beads (part number GB01)

Instructions

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- 1. Pour overnight bacterial culture into a microcentrifuge tube.
- 2. Centrifuge culture to yield a cell pellet (2000g for one minute).
- 3. Completely aspirate the supernatant liquid. Place tube on ice.
- 4. Inspect the volume of the pellet. It should be 300µL or less in order to get efficient homogenization.
- 5. Add volume of zirconium oxide beads equal to the size of the cell pellet.
- 6. Add 0.1mL to 0.6mL buffer (2 volumes of buffer for every volume of cells).
- 7. Snap the caps centrifuge tubes closed.
- 8. Place tubes into the Bullet Blender[®].
- 9. Set controls for SPEED 8 and TIME to 3 minutes. Press Start.
- 10. After the run, remove tubes from the instrument.
- 11. Visually inspect samples. If homogenization is unsatisfactory, run for another two minutes at the SPEED 8.
- 12. Proceed with your downstream application.

SAFETY NOTE!!!

When using a centrifuge to separate your homogenate from the debris and beads, make sure your tubes are balanced.

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