

Bullet Blender® 5 Homogenization Protocol for Adipose Tissue

The protocol described in this document is for the use of the Bullet Blender[®] 5 for the homogenization of fat/adipose tissue (from a variety of animals). If you have difficulty with this protocol, cutting your tissue into smaller pieces will help. This protocol does not specify a particular buffer - you may choose which is most appropriate for your downstream application (nucleic acid isolation, protein extraction, etc.).

Materials Required: Adipose tissue, Bullet Blender[®] 5, homogenization buffer,

pipettor, 5mL Axygen® brand tubes, and 2.0 mm zirconium oxide

beads (part number ZrOB20).

Instructions

1. Cut adipose into appropriately sized pieces for analysis (0.1g - 1g).

- **2. OPTIONAL:** If desired, wash the tissue 3x with 5mL PBS to remove blood and other contaminants from the tissue.
- **3.** Place sample in 5ML tube and add beads to the tube. Use a volume of beads equal to the mass of tissue. **NOTE:** $100 \text{mg} \cong 100 \mu\text{L}$.
- **4.** Add 0.2mL to 2.0mL buffer (2 volumes of buffer for every volume of sample).
- **5.** *Tightly* screw the centrifuge tubes closed and place them into the Bullet Blender[®].
- **6.** Set controls for **SPEED 8** and **TIME 3** minutes. Press start.
- **7.** After the run, remove the tubes from the instrument.
- **8.** Visually inspect samples, if homogenization is unsatisfactory, run for another two minutes at **SPEED 9**
- **9.** Proceed with your downstream application.

SAFETY NOTE!!!

When using a centrifuge to separate your homogenate from the debris and beads, make sure your tubes are balanced.

Date 05/06/2011



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