Bullet Blender[®] 50 Homogenization Protocol for Bladder

The protocol described in this document is for the use of the Bullet Blender[®] 50 for the homogenization of Bladder Tissue. This protocol does not specify a particular buffer - you may choose which is most appropriate for your downstream application (nucleic acid isolation, protein extraction, etc.).

Materials Required:

bladder , Bullet Blender[®] 50, homogenization buffer, pipettor, 50mL skirted centrifuge tubes (Axygen[®] or Corning[®] brand), 4.8mm stainless steel beads (part number SSB48).

Instructions

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- **1.** Cut bladder into appropriately sized pieces for analysis (0.1g 3.5g) and place into a 50mL centrifuge tube.
- **2. OPTIONAL:** If desired, wash the tissue 3x with 5mL PBS to remove blood and other contaminants from the tissue.
- **3.** Add a mass of stainless steel beads (4.8mm) to the tube equal to approximately 6x the mass of your sample.
- 4. Add 0.2 mL to 7mL buffer (2 volumes of buffer for every mass of sample).
- 5. Screw caps onto centrifuge tubes **TIGHTLY**.
- **6.** Place tubes into the Bullet Blender[®] 50.
- 7. Set controls for SPEED 10 and TIME 12 minutes.
- **8.** Remove tubes from the instrument.
- **9.** Visually inspect samples, if homogenization is unsatisfactory, run for another six minutes at the **SPEED 10**.
- **10.** Proceed with your downstream application.

SAFETY NOTE!!!

When using a centrifuge to separate your homogenate from the debris and beads, make sure your tubes are balanced.

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